

RESEARCH ARTICLE

[\(Open Access\)](#)

Isolation, Prevalence and Antimicrobial Resistance of *Salmonella* Strains in Albania from Carcasses in Layers, Turkeys and Ducks

SONILA ÇOÇOLI^{1*}, TANA KIKI¹, JONIDA BOCI², MANJOLA KULIÇI³, NIKOLA PUVACA⁴, PRANVERA CABELI¹, NATALIA SHOSHI¹ AND ANTONIO CAMARDA⁵

¹Veterinary Public Health Department, Faculty of Veterinary Medicine, Agricultural University of Tirana, Rruga: “Pajsi Vodica”, Kodër-Kamëz, Tirana, Albania

²Food Safety and Veterinary Institute, Rruga: “Aleksandër Moisiu” 82, Tirana, Albania

³Food and Biotechnology Department, Faculty of Biotechnology and Food, Agricultural University of Tirana, Rruga: “Pajsi Vodica”, Kodër-Kamëz, Tirana, Albania

⁴Department of Engineering Management in Biotechnology, University Business Academy, Novi Sad, Cvečarska 2, 20100 Novi Sad, Serbia

⁵Department of Veterinary Medicine, University of Bari “Aldo Moro”, Sp. per Casamassima Km 3, 70010 Valenzano, Bari, Italy

*Corresponding author; E-mail: scocoli@ubt.edu.al

Abstract

Salmonellosis is a frontrunner in food-borne diseases with emerging public health concerns. Specific species of *Salmonella* genus present an important poultry and human pathogen. The present study was undertaken in the aim of prevalence detection and antibiogram results estimation in relation to clinical findings in two poultry and one waterfowl species. *Salmonella* spp. were isolated from 15 (19.4%) of 77 examined poultry carcasses samples. High prevalence of Salmonellosis infection was encountered in turkeys (53.33%), followed by ducks (26, 67%) and layers (20%). All *Salmonella* isolates were tested for their susceptibility to 8 selected antimicrobial agents Neomycin (N 30 mcg), Florfenicol (FFC 30 mcg), Amoxicillin (AX 30 mcg), Oxytetracycline (OTC 30 mcg), Norfloxacin (NOR 30 mcg), Enrofloxacin (ENR 10 mcg), Doxycycline (DO 30 mcg) and Trimethoprim-Sulfamethoxazole (SXT 25 mcg) by the agar diffusion method. The overall resistance level was valued as 81, 67%. Out of 15 *Salmonella* isolates, 4 (26, 67%) showed multiple resistance to three or more different tested antimicrobials. Resistance levels in ducks were emerging (100%) toward all of the tested molecules. Emerging high levels of antibiotic-resistance in *Salmonella* isolates ranged from 100% to 46,67% (respectively: Neomycin 100%, Doxycycline 100%, Amoxycillin 100%, Oxytetracycline 100%, Trimethoprim – Sulfamethoxazole 100%, Neomycin 80%, Florfenicol 80%, Enrofloxacin 46, 67% and Norfloxacin 46,67%). The results of the conducted research presented higher prevalence of Salmonellosis in free-range breded poultry (turkeys and ducks) and high antimicrobial rates in ducks. On the other hand, the prevalence of Salmonellosis in layers still remains a concern of food-borne pathogens in food safety chain. Consequently, this study is vital to reveal infection persistence and drug resistance characteristics of *Salmonella* spp. from all of the poultry species which will lay the foundation for follow-up studies of infection persistence and drug resistance mechanisms spread in farm and off-farm environment level.

Keywords: Keywords: *Salmonella* spp.; antibiotic-resistance; poultry; ducks; Albania.

1. Introduction

Salmonellosis is one of the most important bacterial zoonotic diseases, where more than 21 million cases of human Salmonellosis are reported globally in one year. *Salmonellae* live in the gastrointestinal tracts of domestic and wild animals [3] and they are widely

distributed in the environment. Cases of Salmonellosis outbreaks are often related to the consumption of food of animal origin, mainly poultry products, such as eggs and raw chicken [6]. Twenty percent of world poultry products are contaminated with *Salmonella*,

and they can resist for a long time in the animal and human environment [7].

Antimicrobial substances and their resistance has become a problem due to the misuse or overuse of antibiotics, both in human medicine and animal production. This worldwide concern of drug resistance is increased by the fact that a wide variety of antibiotics are not only used for medical and veterinary purposes but also to promote the growth of livestock [2]. Many studies has shown the implication of *Salmonella* in drug resistance and a recent report has shown that tetra and penta-resistance are found in *Salmonella* [8].

This antibiotic-resistant *Salmonella* is a great concern because through the consumption of contaminated food of animal origin will result in health risk to humans [1], [4].

The present study was undertaken in the aim of prevalence detection and antibiogram results estimation in relation to clinical findings in 77 carcasses presented at The Food Safety and Veterinary Institute. The samples analysed were collected from layers, turkeys and ducks.

2. Material and Methods

This study was conducted in Albania, on 77 poultry carcasses during the period of time 2021-2022. After presented at the laboratory, they were examined for post-mortem findings. The most common signs were bronzed enlarged liver with small necrotic foci, anemia and also engorgement of kidneys and spleen.

For the isolation of *Salmonella* spp., 25 g of sample (thigh, breast, wings and liver) from each carcass was

cut into small pieces and blended for 2 min in sterile stomacher bags containing 225 ml of buffered peptone water (0.1%) as a pre-enrichment broth and incubated at 37°C for 24 h. After incubation, 0.1 ml of pre-enrichment culture was transferred into sterile tubes containing 10 ml of Rappaport Vassiliadis broth and the tubes were then incubated at 43°C for 24 h. Thereafter, a loopful of each incubated tube was cultured on Xylose Lysine Desoxycholate agar plates and incubated for 24 h at 35°C. Typical colony of *Salmonella* appears as pink colonies with or without black centers.

After the identification by microscopical examination with Gram stain was carried out the the biochemical test with API 20 E.

All *Salmonella* isolates were tested for their susceptibility to 8 selected antimicrobial agents Neomycin (N 30 mcg), Florfenicol (FFC 30 mcg), Amoxicillin (AX 30 mcg), Oxytetracycline (OTC 30 mcg), Norfloxacin (NOR 30 mcg), Enrofloxacin (ENR 10 mcg), Doxycycline (DO 30 mcg) and Trimethoprim-Sulfamethoxazole (SXT 25 mcg) by the agar diffusion method.

3. Results and Discussion

During this study, from 77 different poultry carcasses tested, 15 cases of *Salmonella* spp. in total were isolated. Eight strains of *Salmonella* spp. were isolated in turkeys, with a high prevalence of 53,33 %, three strains in layers (20%) and four strains in ducks (26,67%).

Table 1. Isolated cases of *Salmonella* spp. from different poultry species

<i>Poultry species</i>	<i>Salmonella spp. positive</i>	<i>Prevalence of Salmonella spp.</i>
Turkeys	8 strains	53.33%
Layers	3 strains	20%
Ducks	4 starains	26.67%

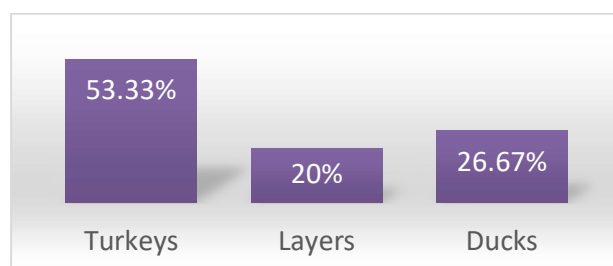


Figure 1. Prevalence of *Salmonella* spp. for each poultry species

All *Salmonella* isolates were tested for their susceptibility to 8 selected antimicrobial agents: Neomycin (N 30 mcg), Florfenicol (FFC 30 mcg), Amoxicillin (AX 30 mcg), Oxytetracycline (OTC 30 mcg), Norfloxacin (NOR 30 mcg), Enrofloxacin (ENR 10 mcg), Doxycycline (DO 30 mcg) and

Trimethoprim-Sulfamethoxazole (SXT 25 mcg) by the agar diffusion method.

The overall resistance level was valued as 81, 67%. Out of 15 *Salmonella* isolates, 4 (26, 67%) showed multiple resistance to three or more different tested antimicrobials. Resistance levels in ducks were emerging (100%) toward all of the tested molecules.

Table 2. Total antimicrobial resistance for 15 *Salmonella* spp. tested strains

<i>Tested antibiotic</i>	<i>Percentage of stains resistance</i>
Neomycin	80%
Flofenicol	80%
Amoxicillin	100%
Oxytetracycline	100%
Norfloxacin	46,67%
Enrofloxacin	46,67%
Doxycycline	100%
TMPX	100%

Emerging high levels of antibiotic-resistance in *Salmonella* isolates ranged from 100% to 46,67% (respectively: Neomycin 100%, Doxycycline 100%, Amoxicillin 100%, Oxytetracycline 100%,

Trimethoprim – Sulfamethoxazole 100%, Neomycin 80%, Florfenicol 80%, Enrofloxacin 46, 67% and Norfloxacin 46,67%).

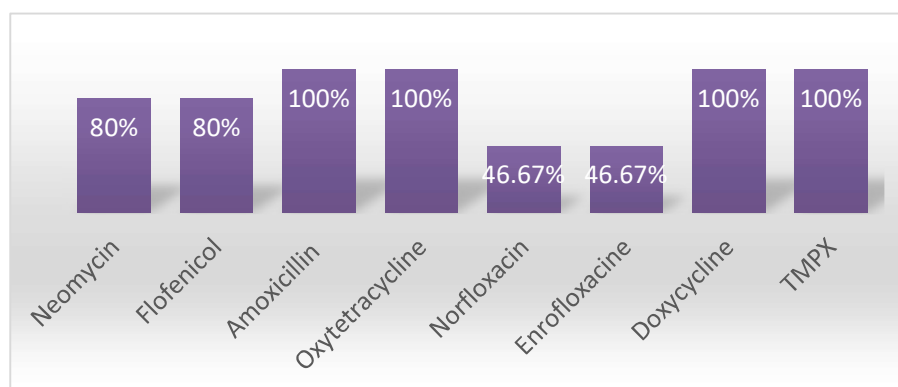


Figure 2. Total AMR for 15 *Salmonella* spp. tested strains

The results obtained coincide with other studies conducted in this field. Increased prevalence of antimicrobial-resistant *Salmonella* presents a severe risk to human health. Transmission of antimicrobial resistance genes (ARGs) to the environment and eventually humans further aggravates the situation [5].

4. Conclusions

Salmonella spp. were isolated from 15 (19.4%) of 77 examined poultry carcasses samples. High prevalence of Salmonellosis infection was encountered in turkeys (53,33%), followed by ducks (26, 67%) and layers (20%), where Salmonellosis in layers still remains a concern of food-borne pathogens in food safety chain

The results of the conducted research presented higher prevalence of Salmonellosis in free-range breded poultry (turkeys and ducks) and high antimicrobial rates in ducks (100 %). Out of 15 *Salmonella* isolates, 4 (26, 67%) showed multiple resistance to three or more different tested antimicrobials. The overall resistance level was valued as 81, 67%.

These findings highlight the necessity to reduce the pathogen prevalence through continuous control of its presence in animals and environment and also the need for government entities, researchers and poultry producers to find ways to reduce the impact of antibiotic use in poultry, focusing especially on active

surveillance of multi drug resistance strains and finding alternatives to substitute antibiotics.

5. References

1. Barreto M., Castillo-Ruiz M., Retamal P. ***Salmonella enterica* a review or the trilogy agent, host and environment and its importance in Chile.** Rev. Chil. Infectología. 2016;33(5):547–557.
2. Davies, J.; Davies, D. **Origins and evolution of antibiotic resistance.** Microbiol. Mol. Biol. Rev. 2010, 74, 417–433.
3. De Jong B., Ekdahl K. **The comparative burden of salmonellosis in the European Union member states, associated and candidate countries.** BMC Public Health. 2006;6:1–9. doi: 10.1186/1471-2458-6-4.
4. H.L, Chan K.G, Lee L.H. ***Salmonella*: A review on pathogenesis, epidemiology and antibiotic resistance.** Front. Life. Sci. 2015; 8 (3):284–293.
5. Kumar, D., Pornsukarom, S., Thakur, S. (2019). **Antibiotic Usage in Poultry Production and Antimicrobial-Resistant Salmonella in Poultry.** In: Venkitanarayanan, K., Thakur, S., Ricke, S. (eds) Food Safety in Poultry Meat Production. Food Microbiology and Food Safety. Springer, Cham. https://doi.org/10.1007/978-3-030-05011-5_3
6. Tan, S.J.; Nordin, S.; Esah, E.M.; Mahrer, N. ***Salmonella* spp. in Chicken: Prevalence, Antimicrobial Resistance, and Detection Methods.** Microbiol. Res. 2022, 13, 691–705. <https://doi.org/10.3390/microbiolres13040050>
7. Vestby LK, Moretro T, Langsrud S, Heir E, Nesse LL. **Biofilm forming abilities of Salmonella are correlated with persistence in fish meal-and feed factories.** BMC Vet Res 2009; 5: 20.
8. Y. Xiang, F. Li, N. Dong, S. Tian, H. Zhang, X. Du, X. Zhou, X. Xu, H. Yang, J. Xie, C. Yang, H. Liu, S. Qiu, H. Song, **Investigation of a Salmonellosis outbreak caused by multidrug resistant *Salmonella typhimurium* in China** Front. Microbiol., 11 (2020), p. 801